

Functional Assays of Membrane-Bound Proteins with SAMDI-TOF Mass Spectrometry**

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Proteins that span the lipid bilayer membrane—including cell adhesion molecules, growth factor and cytokine receptors, and transporters—mediate a broad range of interactions of the cell with its environment and are prominent targets in many drug-discovery programs. The difficulties associated with traditional cell-based and micelle-solubilized assays have motivated significant work in the past decade to immobilize proteins within lipid bilayers supported on flat surfaces^[1–7] and has made possible the reconstitution of membrane proteins and assays of their activities by fluorescence and evanescent wave methods.

Our objective in the current work was to use mass spectrometry (MS) to perform functional assays of membrane-resident proteins. Since MS methods do not require labels, they simplify and increase the scope of assays and provide molecular information about the analyte. Our experiment is presented in Figure 1 and begins with solubilization of the seven-transmembrane photoreceptor rhodopsin in a nanodisc. The nanodisc strategy is based on the self-assembly of lipid molecules within a membrane scaffold protein, giving a circular patch of soluble lipid bilayer that can additionally contain a transmembrane protein.^[8–10] We expressed a membrane scaffold protein (MSP) having a hexahistidine (his₆) tag at its N terminus and prepared nanodiscs containing the rhodopsin protein and the lipid 1-palmitoyl-2-oleoyl-*sn*-glycero-3-phosphocholine (POPC) as described in the Supporting Information.^[11–16] A MALDI-TOF mass spectrum of the nanodiscs showed a clear peak at m/z 25.6 kDa for the

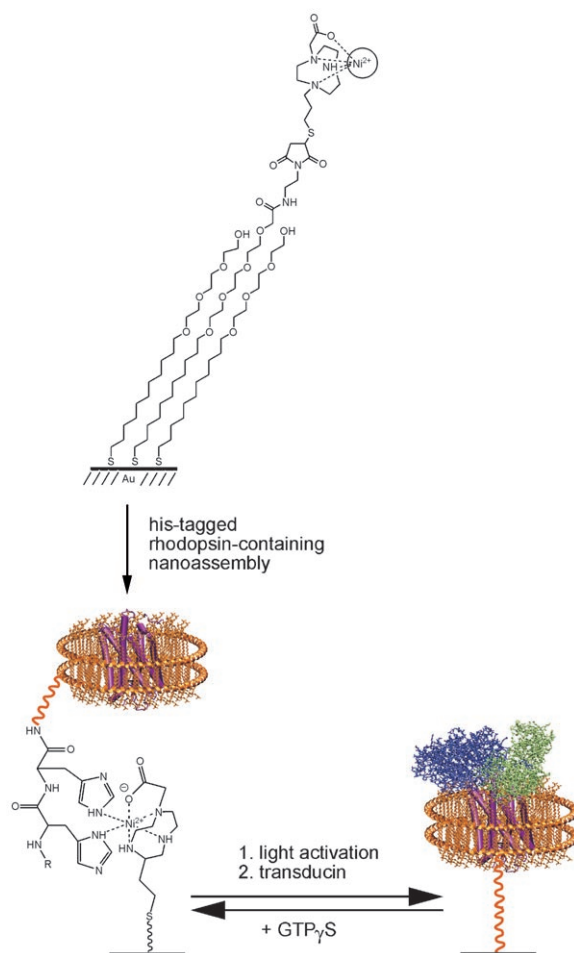


Figure 1. The seven-transmembrane protein rhodopsin is immobilized on a self-assembled monolayer by way of a his-tagged membrane scaffold protein. Upon activation of rhodopsin with light, the receptor binds the transducin complex and can be detected using SAMDI-TOF mass spectrometry. GTP γ S = guanosine 5'-(thiotrisphosphate).

membrane scaffold protein, and at 760.1 Da for the lipid molecule.

We first examined the immobilization of the lipid-loaded nanodiscs to a self-assembled monolayer. To prepare this surface, we immobilized a thiol-terminated triazacyclononane-derived ligand (Actacn), which is functionally analogous to the common nitrilotriacetic acid (NTA) ligand for immobilization of his-tagged proteins,^[17,18] to a monolayer presenting maleimide and tri(ethylene glycol) groups (see the Supporting Information).^[19] The former provides a defined density of attachment sites for the nanodiscs, and the latter are effective at preventing nonspecific interactions of proteins

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with the surface. We prepared nanodisc assemblies containing only the membrane scaffold protein and POPC and monitored the immobilization of this construct using surface plasmon resonance spectroscopy (SPR; Figure 2A). The nanodisc efficiently bound to the Actacn-Ni^{II} complex on

ciently immobilized on self-assembled monolayers and observed by both surface plasmon resonance spectroscopy and SAMDI mass spectrometry.

Nanodiscs containing the protein rhodopsin were subsequently self-assembled at a mol ratio of 700:10:1 POPC/MSP/rhodopsin. Again, SPR and SAMDI MS confirmed the immobilization of the protein-loaded nanodiscs on monolayers. The SAMDI spectrum reveals peaks at $[M]^+$ at m/z 25.6 kDa and $[2M]^+$ at m/z 12.8 kDa for the MSP and a peak at $[M]^+$ at m/z 43.6 kDa for rhodopsin (Figure 3A).

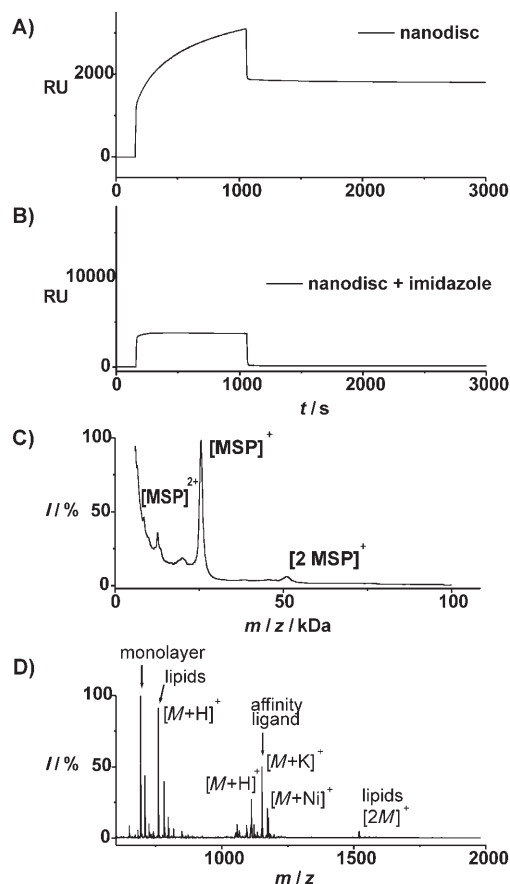


Figure 2. Characterization of his-tagged nanoassemblies containing lipids immobilized onto Actacn-presenting monolayers. SPR binding curve of nanodiscs in the absence (A) and presence (B) of imidazole (150 mM). C,D) SAMDI-TOF mass spectra of duplicate samples to those in A). Sinapinic acid was used as a matrix to observe protein (C) and 2,4,6-trihydroxyacetophenone (THAP) was used to observe small molecules (D). RU=resonance units.

the monolayer and remained bound during a wash with buffer. The addition of imidazole, which complexes the Ni^{II} ion, prevented binding of the his-tagged nanodisc and demonstrates the effectiveness of the oligo(ethylene glycol) groups to prevent unwanted protein adsorption. Following immobilization of the nanodisc, we used a variant of MALDI-TOF mass spectrometry to characterize the surface. This technique, known as self-assembled monolayers for matrix-assisted laser desorption/ionization time-of-flight mass spectrometry (SAMDI),^[20–23] shows a peak at $[M]^+$ m/z 25.6 kDa for the his-tagged membrane scaffold protein (Figure 2C). Another spectrum acquired at lower mass range also revealed peaks at $[M]^+$ at m/z 760.1 and at $[2M]^+$ at m/z 1520.2 that correspond to the lipid molecules of the nanodisc (Figure 2D). Hence, the nanodiscs could be effi-

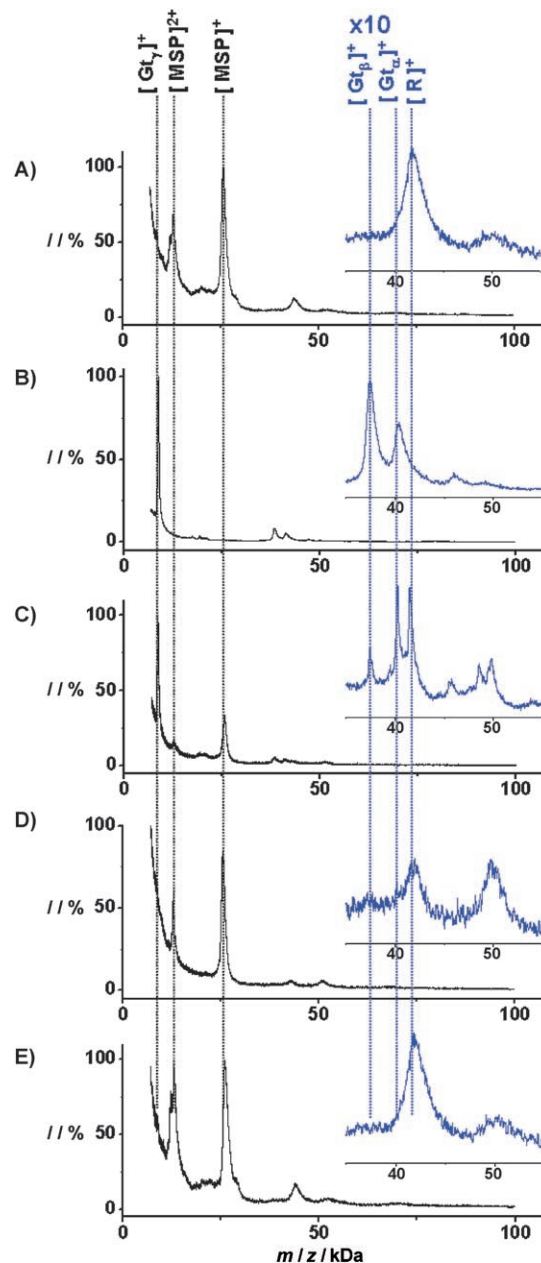


Figure 3. A) SAMDI-TOF mass spectrum of rhodopsin-loaded nanoassemblies immobilized on a monolayer. B) MALDI-TOF mass spectrum of transducin. C) Immobilized rhodopsin exposed to light binds to the transducin protein complex. Peaks for the transducin α (Gt_α), β (Gt_β), and γ (Gt_γ) subunits, rhodopsin (R), and the membrane scaffold protein (MSP) are indicated at the top. D) A control experiment performed in the dark prevented binding of transducin to immobilized rhodopsin, as did inclusion of the nucleotide inhibitor GTP γ S (E).

We then performed a functional assay based on the photoactivation of rhodopsin, whereby photoisomerization of 11-*cis*-retinal, a lysine-bound chromophore, leads to a conformational change that allows for binding of the transducin protein complex.^[24] We incubated the Actacn-presenting monolayers with a mixture of the rhodopsin-loaded nanodiscs and transducin (10 μ M) for 30 min at room temperature and in the field of ambient light. A SAMDI spectrum reveals peaks corresponding to both MSP and transducin (Figure 3C). The peak for the γ subunit of transducin has a greater intensity than those for the α and β subunits and for rhodopsin. We frequently find that lower-molecular-weight proteins yield more intense peaks. Hence, a comparison of peaks for γ -transducin and MSP provides the clearest evidence for activity of the rhodopsin receptor. A control experiment performed in the dark, which prevented the activation of the rhodopsin receptor, was analyzed by SAMDI and revealed peaks corresponding only to the MSP and rhodopsin (Figure 3D). This protein–protein interaction is also regulated by small molecules, including the nonhydrolyzable nucleotide GTP γ S which binds to the α subunit of transducin (Gt $_{\alpha}$) and blocks its interaction with rhodopsin.^[25] We treated a monolayer presenting the rhodopsin–transducin complex with an excess of GTP γ S. Analysis of this substrate by SAMDI showed that the inhibitor led to dissociation of transducin from the nanodisc-bound rhodopsin (Figure 3E).

This example establishes a route to performing functional assays of membrane-bound proteins that are immobilized on biochip surfaces. Several aspects of this approach are significant. Nanodiscs provide a means to control the size of the bilayer,^[11] the lipid composition^[26] and the stoichiometry of the incorporated membrane protein,^[27] permitting these macromolecules to be treated as soluble proteins. The self-assembled monolayers provide well-defined substrates for protein immobilization. The density of the immobilized proteins can be controlled by adjusting the density of the Actacn ligand and therefore a more uniform activity of protein. We note that a range of other immobilization chemistries are available, including the cutinase affinity tag that gives a covalent immobilization of protein.^[28–30]

Finally, the monolayers enable the use of mass spectrometry to directly observe the proteins immobilized to the monolayer and any interacting partners. This strategy obviates the need for fluorescent or radioisotopic labels, provides mass information on the species bound to biochip, and therefore can be effective at identifying (and ignoring) background species, at identifying posttranslational modifications of immobilized proteins, and at performing multi-analyte assays. These attributes bring to membrane-bound proteins the full range of techniques that have been important for assaying the activities of soluble proteins.

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